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Computerization of Virus Data and Its Usefulness in Virus Classification

Summary

Data on 537 Arboviruses and 180 other viruses have been collected and coded in two different formats. These data include information not only regarding the taxonomy and history of isolation, but also regarding the properties of biomacromolecules, proteins and nucleic acids. Information on antigenic relationships, histopathology and experimental viremia is also included. This information is stored in formats which allow the manipulation and analysis of data by dBASE III PLUS and MICRO-IS. A set of programs was written for interconversion and editing purposes. Transmission electron micrographs are scanned and stored. This stored information can be used in viral classification as shown by carrying out analysis of data on the Bunyaviridae family.

Introduction

In recent years advancements in techniques of protein sequencing, nucleic acid sequencing and hybridoma technology have been used by molecular virologists to gain insight into the structure and behavior of viruses. Information generated through such a set of experiments is large and requires storage in a computer-readable form so that access can be gained and the data analyzed. A large number of molecular and epidemiological studies have also been carried out to understand and model the outbreaks of epidemics at the vector and host level. Such modelling studies will have little predic-

tive value if the data on molecular, vector, natural and experimental hosts and from cell culture studies are not used for analysis, suggesting a need for computerization of virus data.

Some of the important groups who organize virus data include: (i) the International Committee on Taxonomy of Viruses (ICTV) [1–3]; (ii) the International Catalogue of Arboviruses [4]; (iii) WHO Center for Collection and Evaluation of Data on Comparative Virology in Munich [personal commun.], and (iv) Virus Data Exchange (VIDE) project of the Australian National University [personal commun.].

To complement these efforts and to aid virologists, molecular biologists, clinical personnel,

epidemiologists, and industrial users, we have initiated the computerization of data on animal viruses. In the first phase of this project, information on arboviruses has been computerized. An open-ended numerical coding system, pictorial data storage and standardization of vocabulary are some of the features of the data bank which are discussed in the following sections. It is pointed out that this data bank can be used as an aid to classify viruses objectively. At present, information on 537 arboviruses has been coded and is available in computer-readable form. Information on another 180 viruses is also computerized.

Coding of Virus Data in dBASE III PLUS

Information on viruses has been coded using the dBASE III PLUS data base management system [5, 6]. The format chosen allows the user to extract all or part of the information on a particular virus. Information is stored under 16 different categories which are listed in table 1. There are several subcategories and they vary in number and organization. In order to extract information based on the properties

Table 1. Categories incorporated for coding data in dBASE III PLUS format

- 1 Virus status and distribution
- 2 The original source of the virus
- 3 Method of isolation and validity
- 4 Physicochemical properties of virus
- 5 Stability of infectivity and virulence
- 6 Virion morphology
- 7 Morphogenesis
- 8 Hemagglutination
- 9 Antigenic relationship
- 10 Susceptibility of cell systems
- 11 Natural host range
- 12 Experimental viremia
- 13 Histopathology
- 14 Human disease
- 15 Links with other data banks
- 16 References

of viruses, it is essential to make use of a controlled vocabulary during coding and querying. A controlled vocabulary is essential since the information developer also uses synonyms and acronyms in the report. Examples of some of the controlled vocabulary terms are given in table 2. We have developed a dictionary of

Table 2.

Words and their possible synonyms incorporated in the data bank A single word and its possible synonyms

Idoxuridine: 5-iodo-2-deoxyuridine, Dendrid, Herpid, Kerecid, Stoxil

Idoxuridine = An antiviral agent

5-iodo-2-deoxyuridine = The chemical name of idoxuridine

Dendrid = Trade name of idoxuridine eye drops

Kerecid = Trade name of idoxuridine eye drops

Herpid = Trade name of 5% idoxuridine

An acronym and its full form

SAM = S-Adenosyl-L-methionine

SAM = An intracellular carrier and donor of activated methyl groups

A virus and its possible synonyms

Japanese encephalitis virus = Japanese B virus, Russian autumn encephalitis virus

An arthropod and its possible synonyms

Culex quinquefasciatus = Culex fatigans, Culex (Culex) fatigans, C. fatigans

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Fig. 1. A sample dBASE data file for Japanese encephalitis virus.

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VIRUS STATUS AND DISTRIBUTION
                                                          STABILITY OF INFECTIVITY AND VIRULENCE
  Accession no.: A0195-000
                                                            Effect of ether: yes
  Virus name, prototype : Japanese Encephalitis
                                                            Effect of chloroform : not known
                                                            Effect of deoxycholate : yes
                          (Nakayama)
  Abbreviation: JBE
  Antigenic group: B
                                                          VIRION MORPHOLOGY
  Arbovirus status : ARBOVIRUS
                                                            Shape of the virus is : SPHERICAL
                                                            Dimensions of the virus: 48.6-57.6 nm
  Family : FLAVIVIRIDAE
  Genus : FLAVIVIRUS
                                                            Method of measurement : EM
  Taxonomic status : FLAVIVIRUS
                                                            Nature of envelope/surface projections : SPIKES
                                                            Nucleocapsid dimensions: 27.3-32.3 nm
  Biosafety level (SALS rating): 3
  Geographical distribution:
          1) INDIA
                                                          MORPHOGENESIS
          2) JAPAN
                                                            Site of virion assembly in cell:
          3) JAVA
                                                                CYTOPLASM
          4) KOREA
                                                            Site of virion accumulation in cell:
          5) NORTH AND S.E. ASIA
                                                                CYTOPLASM VACUOLUM
          6) NORTHEASTERN ASIA
          7) TAIWAN
                                                          HEMAGGLUTINATION
          8) THAILAND
                                                           Hemagglutination was observed
                                                           Antigenic source for testing hemagglutination :
                                                               SMB.EXT.BY BORATE-KCl, pH 9.0; ACETONE-ETHER;
ORIGINAL SOURCE OF THE VIRUS
                                                               SUCROSE-ACETON
The original source of the virus was isolated by :
                                                           Erythrocytes used for hemagglutination: GOOSE
      T.Mitamura and M.Kitaoka, et al.
The place of isolation of the original source was:
                                                           Minimum pH for hemagglutination: 6.0
                                                           Maximum pH for hemagglutination: 7.2
      Tokyo,
                                                           Optimum pH for hemagglutination: 6.6
The country of isolation of the original source
                                                           Minimum temperature for hemagglutination: 4 degC
was : Japan
The animal from which the original source was
                                                           Maximum temperature for hemagglutination: 37 degC
                                                           Optimum temperature for hemagglutination: 25 degC
isolated : MAN
The age of the source animal is: 19 YEARS
                                                           Serological methods recommended : HI, CF, NT
The sex of the source animal is : male
The virus was isolated from the sample : BRAIN
                                                          ANTIGENIC RELATIONSHIP
The signs of illness during original isolation
                                                               Related antigenically to other flaviviruses
were : ENCEPHALITIS
The sample collection date is (D/M/Y): 21/08/1935
                                                               (Group B)
The sample collection method is : AUTOPSY
                                                               Viruses antigenically related: West Nile,
The sample collection place is:
                                                               Murrey Valley Encephalitis, Saint Louis
                                                               Encephalitis.
HOSPITAL IN TOKYO, JAPAN
The macrohabitat is: URBAN
                                                          SUSCEPTIBILITY OF CELL SYSTEMS
Method of storage until inoculated: NO STORAGE
                                                                                1) CHICK EMBRYO
METHOD OF ISOLATION AND VALIDITY
                                                              Cell type
                                                                  Cell line or prim.cul.: PC
Inoculation date is (D/M/Y): 21/08/35
The animal inoculated: WN MICE
                                                                  CPE Observed : YES
The route of inoculation is: INTRACEREBRAL
                                                                  Plaque observed : YES
Reisolation of the virus was attempted.
                                                                  Day Plaque observed: 5
Homologous antibodies were not produced by source
                                                              Cell type
                                                                                2) HUMAN EPITHELIAL
                                                                  Cell line or prim.cul.: CL
animal.
                                                                  CPE Observed : NO
Tests used: NT
                                                                  Plaque observed : NOT KNOWN
PHYSICOCHEMICAL PROPERTIES
                                                                               3) L MOUSE CELLS
                                                              Cell type
                                                                  Cell line or prim.cul.:
The virus is an RNA virus
The virus is not a DNA virus
                                                                  CPE Observed: NO
The virus is a single stranded virus
                                                                  Plaque observed : NOT KNOWN
The virus is not a double stranded virus
                                                              Cell type
                                                                               4) VERO
                                                                  Cell line or prim.cul.: CL
No. of pieces of the genome are 1
Number of virion polypeptides 3
                                                                  Virus passage history: P-52
Details of virion polypeptides
                                                                  CPE Observed : NOT KNOWN
     9,000-10,000 (MEMBRANE POLYPEPTIDE),
                                                                  Plague observed : YES
     13,000-15,000 MW (CAPSID POLYPEPTIDE)
                                                                  Day Plaque observed: 4
Virion density in (gm/cm-cube) 1.19
                                                                  Size of Plaques: 3.0 mm
Virion density measured in SUCROSE
                                                                  Titer (Plaque): 9.1 (PFU/ml)
Virion sedimentation coefficient (in s) 200
                                                              Cell type
                                                                              5) LLC-MK2
                                                                  Cell line or prim.cul.: CL
Nucleocapsid density (in gm/cm-cube) 1.31
                                                                  CPE Observed : NOT KNOWN
Nucleocapsid density measured in SUCROSE
                                                                  Plaque observed : YES
                                                                  Day Plaque observed: 3
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Size of Plaques: 8.0 mm Titer (Plaque): 9.4 (PFU/ml) Cell type 6) C6/36 Cell line or prim.cul.: CL CPE Observed: NOT KNOWN Plaque observed : YES Cell type 7) PS Cell line or prim.cul.: CL Virus passage history: P-43 CPE Observed : YES Day CPE observed: 3 Extent of CPE: 100 % Titer (CPE): 8.0 TCD50/ml Plague observed: YES Day Plaque observed: 3 Size of Plaques: 5.0 mm Titer (Plaque): 8.3 (PFU/ml) NATURAL HOST RANGE Species : ANOPHELES BARBIROSTRIS No. of isolations: 1 Species : CULEX TRITAENIORHYNCHUS Country from which species collected: INDIA No. of isolations: 1 Species : ANOPHELES HYRCANUS Country from which species collected: INDIA No. of isolations: 1 Species : MAN Country from which species collected: INDIA No. of isolations: MANY Species : CULEX PSEUDOVISHNUI Country from which species collected: INDIA No. of isolations: 1 Species : CULEX FUSCOCEPHALA Country from which species collected: INDIA No. of isolations: 1 Species : HORSES Country from which species collected : JAPAN No. of isolations : MANY Species : BIRDS Country from which species collected: JAPAN No. of isolations: MANY Species : CULEX PIPIENS Country from which species collected : JAPAN Species : CULEX GELIDUS Country from which species collected : JAPAN Species : CULEX VISHNUI Country from which species collected : JAPAN No. of isolations: 1 Species : MAN Country from which species collected : JAPAN

No. of isolations: 3 Species : POIKILOTHERMIC HOSTS Country from which species collected : KOREA Species : CX.PIPIENS Country from which species collected: NORTH AND S.E. ASIA Species : CX.TRITAENIORHYNCHUS Country from which species collected: NORTHEASTERN ASIA No. of isolations : MANY Species : CX.GELIDUS Country from which species collected: S.E. ASIA Species : CX.ANNULUS Country from which species collected: TAIWAN Species : BATS Country from which species collected: TAIWAN Species : POIKILOTHERMIC HOSTS Country from which species collected : TAIWAN Species : CX.FUSCOCEPHALA Country from which species collected: THAILAND EXPERIMENTAL VIREMIA 1) Experimental animal used : MICE Age of the animal: NB Route of inoculation : IC Inoculation dose: 0.01 Evidence of infection : DEATH Average Survival Time (days) from : 3 Average Survival Time (days) upto: 4 Titer (log10/ml) : 8.0 2) Experimental animal used : MICE Age of the animal: NB Route of inoculation : IP Inoculation dose: 0.01 Evidence of infection : DEATH Average Survival Time (days) from: 4 Average Survival Time (days) upto : 5 Titer (log10/ml): 8.0

3) Experimental animal used: MICE
Age of the animal: WN
Route of inoculation: IC
Inoculation dose: 0.03
Evidence of infection: DEATH
Average Survival Time (days) from: 5
Average Survival Time (days) upto: 6

Titer (log10/ml): 7.0

4) Experimental animal used: MICE
Age of the animal: WN
Route of inoculation: IP
Inoculation dose: 0.2
Evidence of infection: DEATH
Average Survival Time (days) from: 6
Average Survival Time (days) upto: 10
Titer (log10/ml): 1.0

No. of isolations : MANY

Species : CX.TRITAENIORHYNCHUS

Country from which species collected : JAVA

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5) Experimental animal used: MONKEYS
                                                                      CNS PLEOCYTOSIS
              Route of inoculation : IC
                                                                Category of human disease:
              Inoculation dose • 0.2
                                                                      ENCEPHALITIS
              Evidence of infection: DEATH
                                                              LINKS WITH OTHER DATA BANKS
            6) Experimental animal used: HAMSTER
              Route of inoculation : IC
                                                              ID CODE as in EMBL :
              Evidence of infection : DEATH
                                                              M62934; M18370; M55506; M73710; D00037; N00037;
                                                              D90195; M14933; M15337; M16574; D90194;
            7) Experimental animal used : GUINEA PIG
              Route of inoculation : IC
                                                              ID CODE as in GENBANK:
              Evidence of infection: VIREMIA
                                                             M62934; M14933; M15337; M18370; M55506; M73710; M16574; D90195; D90194; D00037; N00037;
           8) Experimental animal used: RABBIT
              Route of inoculation : IC
                                                              ID CODE as in NBRF :
              Evidence of infection: VIREMIA
                                                              A27403\ A26465; A27844;
           9) Experimental animal used : CHICK
                                                              ID CODE as in HDB :
              Route of inoculation: IC
                                                              1915; 1916; 1917; 1918; 1919; 1920; 1921; 1922;
                                                             1923; 1924; 1925; 1926; 1927; 1928; 1929; 1939;
1931; 1932; 1933; 1934; 1935; 1936; 1937; 1938;
              Evidence of infection: VIREMIA
          10) Experimental animal used : HORSES
                                                              1939;
             Route of inoculation : SC
             Evidence of infection: VIREMIA
                                                             REFERENCES
                                                              1)Mitamura, T., et al. 1935. Kansai Iji
          11) Experimental animal used: BATS
                                                             260-261:1-5.
             Route of inoculation : SC
                                                             2)Mitamura, T., et al. 1936. Trans. Soc. Path. Jap.
             Evidence of infection: VIREMIA
                                                             26:429-452.
                                                             3)Mitamura, T., et al. 1938. Tokyo Ijishinshi
          12) Experimental animal used:
                                                             3076:766-777.
             EMBRYONATED EGGS
                                                             4) Mitamura, T., et al. 1936. Ibid. 3006:3149-3156.
                                                             5)Mitamura, T., et al. 1936. Ibid. 3006:3157-3161.
             Route of inoculation: YS
             Evidence of infection: DEATH
                                                             6)Mitamura, T., et al. 1938. Ibid. 3076:771-773.
                                                             7) Mitamura, T., et al. 1937. Trans. Soc. Path. Jap.
                                                             27:573-580.
  Animal on which lesions observed and characters
                                                             8) Mitamura, T., et al. 1938. Ibid. 28:135-145.
  of lesions:
                                                             9)Mitamura, T., et al. 1939. Ibid. 29:92-105.
          1) MAN :
                                                             10)Mitamura, T., et al. 1940. et al. 1940. Ibid.
             ENCEPHALITIS
                                                             30:561-570.
          2) Animal not defined:
                                                             11) Mitamura, T., et al. 1938. Tokyo Ijishinshi
             ACUTE ENCEPHALITIS
                                                             3076:779-789.
  Organs and tissues effected in animal:
                                                             12)Mitamura, T., et al. 1936. Tokyo Ijishinshi
          1) Animal not defined:
                                                             3006:3162-3169.
             BRAIN
                                                             13) Mitamura, T., et al. 1936. Ibid. 3006:3170-3172.
          2) MAMMAL :
                                                             14) Mitamura, T., et al. 1937. Ibid. 3030:1145-1155.
                                                             15)Mitamura, T., et al. 1938. Ibid. 3030:778-779.
16)Mitamura, T., et al. 1938. Ibid. 3079:1097-1139.
             BRAIN
          3) LOWER VERTEBRATES:
             BRAIN
                                                             17)Clarke, D.H. and Casals, J. 1958.
  Category of tropism in animal:
                                                             Am.J.Trop.Med.Hyg. 7:561-573.
          1) Animal not defined:
                                                             18)Intl.Cata.of Arboviruses, 1985.pp. 511-512.
             NEUROTROPIC, ENCEPHALITIS
                                                             19) Gresser, I., et al. 1958. Am. J. Trop. Med. Hyg.
                                                             7:365-373.
HUMAN DISEASE
                                                             20)Gresser, I., et al. 1958. Jsp.J.Trop.Med.
  Human disease in nature is significant
                                                             28:243-248.
  Human disease leading to death is significant
                                                             21)Buescher, E.L., et al. 1959.
 Human disease is significantly residual
                                                             J.Immunol.83:582-626.
 Laboratory infection is reported to be
                                                             22)Buescher, E.L., et al. 1962.
 subclinical
                                                             Am.J.Vet.Res.23:1157-1163.
 Laboratory infection is reported to be an overt
                                                             23)Gould, D.J., et al. 1962.
 disease
                                                             Trans.Roy.Soc.Trop.Med.Hyg.56:429-435.
 Clinical manifestations:
                                                             24) Scherer, W.F., et al. 1959. Am.J. Trop. Med. Hyg.
        FEVER
                                                             8:644-722.
        HEADACHE
                                                             25)Rivers, T.M. and Horsfall, F.L. 1959.Viral and
        PROSTRATION
                                                             Rickettsial Infections of Man 3rd Ed. pp. 312-319.
        STIFF NECK
                                                             26) Theiler, M. 1957.
        CNS SIGNS
                                                             Proc.Soc.Exp.Biol.Med.96:380-382.
        ENCEPHALITIS
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synonyms and acronyms. At present 1,400 words are included in this dictionary. We are also making use of those controlled vocabulary terms which are already used by hybridoma data bank developers [7] and protein sequence data bank developers [8]. A sample dBASE data file for the Japanese encephalitis virus of Flaviviridae family is given in figure 1. It can be seen from figure 1 that we have not fed sequences of either nucleic acids or proteins of the virus, but pointers are available through accession numbers given in EMBL nucleotide sequence data banks and Genbank for nucleic acid sequences. Protein sequence information can be obtained using NBRF-PIR accession numbers. Information on the 3D structure of the proteins can also be obtained from the Protein Data Bank because in the NBRF PIR sequence data bank there is a field which indicates whether 3D data are available in the Protein Data Bank of Brookhaven National Laboratory in the USA. Information regarding monoclonals and hybridomas is not fed into our data bank but accession numbers in the Hybridoma Data Bank (HDB) of IUIS-CO-DATA are provided. It may be pointed out that HDB is available on Microbial Strain Data Network (MSDN) through Telecom Gold in the UK and Dialcom in the USA. The protein and nucleic acid sequence data banks are also available online through various USA and European networks. Thus, wherever possible pointers to gain access to computerized data are provided:

Pictorial Data Storage

Many times in biology, pictorial information reveals important features which are difficult to point out by coding the information in the alpha-numeric form. Electron microscopic pictures of biological systems provide very high resolution information regarding the structure and organization at the micro level. This is particularly true for transmission electron micrographs at high resolution which reveal the morphology of the virus. Several methods in electron microscopy, such as metal shadowing, negative staining and freeze fracture, have been employed to study the structure of viruses [9]. The three-dimensional structures of viruses can be produced using computer analysis and can also be determined by fast Fourier reconstruction [10]. We have therefore stored transmission electron micrograph images of viruses on the computer in the TIFF format.

Development of Numerical Codes

The hard data on viruses can be divided into two parts: attributes, and measurable quantities. Comparison of the measurable quantities is possible if the input data are standardized and then coded. In order to achieve this we have developed a new numerical code to describe virus properties. Our data on viruses were divided into three main categories: (a) binary; (b) numeric, and (c) character information, as has been done in the RKC codes for microbes [11]. The codes for some of the virus properties are given in table 3. Data coding on the size of virus, virion morphology, etc., is carried out by using codes which support numeric data, while the information on the history and distribution of the viruses is coded using codes which support character data. The coding system is open ended and makes it possible to update and add new codes from time to time. The addition of new codes does not alter the structure of the data bank. This is mainly because we have distinguished between the experiments giving negative results and experiments not carried out to check the properties.

The coded information is entered in the computer to form a flat file having a large

Table 3. RKC-like codes for some of the virus properties

Codes of binary data

007564 Storage of the virus after isolation is at room temperature (25°)

007565 Storage of the virus after isolation is from 4° to room temperature (25°)

007566 Storage of the virus after isolation at 4°

007567 Storage of the virus after isolation from -20 to 4°

007568 Storage of the virus after isolation at -20°

007569 Storage of the virus after isolation from -20 to -70°

007570 Storage of the virus after isolation at -70°

014201 Type of nucleic acid is DNA

014202 Type of nucleic acid is RNA

014203 Strandedness of nucleic acid is single

014204 Strandedness of nucleic acid is double

014205 Polarity of the nucleic acid is positive

014206 Polarity of the nucleic acid is negative

020201 Mode of transmission of the virus is vertical

020202 Mode of transmission of the virus is horizontal

020203 Mode of transmission of the virus is mechanical

Codes for numeric data

532101 Molecular weight of the entire genome

532103 Percentage weight of the nucleic acid

532106 Percentage composition of base A in entire genome

532107 Percentage composition of base T in entire genome

532108 Percentage composition of base G in entire genome

532109 Percentage composition of base C in entire genome

532110 Percentage composition of base U in entire genome

532315 Percentage weight of the lipids

542101 Virion density in grams per cubic centimeter

542102 Virion sedimentation coefficient in S

Codes for character data:

c07415 Place of collection of the animal from which virus was first isolated

c07418 Macrohabitat of the place of collection of the animal from which virus was first isolated

c07419 Microhabitat of the place of collection of the animal from which virus was first isolated

c14322 Antigenic group of the virus

c14323 Taxonomic status of the virus

number of fields, each field having variable length. Such a flat file structure is adapted mainly because the data base management system called MICRO-IS, developed at the National Institutes of Health, Bethesda, Md. [12], to carry out analysis of microbial data, can be used.

A set of programs is written which converts properties expressed in words into numerical code. Such a software package to automatically convert the dBASE III PLUS data file into MICRO-IS input format is available.

Identification and Classification of Viruses

An example is given here to classify viruses from the family of Bunyaviridae using the sys-

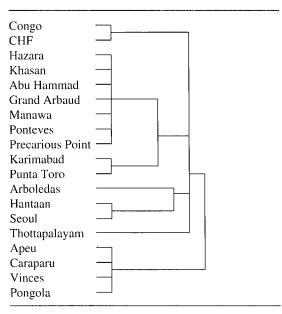


Fig. 2. A possible classification of viruses among the family Bunyaviridae. Nairoviruses, Congo and Crimean hemorrhagic fever (CHF) form one cluster. Nairoviruses, Hazara, Khazan and Abu Hammad, are part of the cluster formed by Uukuvirus, Grand Arbaud, Manawa, Ponteves, and Precarious Point. The Phleboviruses, Karimabad and Punto Toro, are closest to Uukuviruses thus supporting recent conclusions regarding the evolutionary relationship between these two genera [13].

tem. Four properties from 19 randomly chosen members of the Bunyaviridae family are used to create a weight matrix. These are: (i) type of vector; (ii) type of host; (iii) information about the genome, and (iv) results from hemagglutination experiments. Single linkage cluster analysis was carried out using the above-mentioned weight matrix. Results given in figure 2 point out that the weight matrix formed by only the above four properties does give results which are very similar to what we know today about the classification of viruses from the Bunyaviridae family. A better resolution can be achieved if a weight matrix is formed using many properties which are listed in our data bank.

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Availability of Data

These virus data are available online to all the MSDN users. Access to MSDN can be gained not only through Telecom Gold and Dialcom but also through Internet. The software being used to gain access to and analyze information is called INFO. The data are actually situated at Base de Dados Tropical, Campinas, Brazil. At present only alphanumeric data are available online. We will also provide a copy of the data base on floppy diskette at nominal handling charges.

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References

- I Matthews REF: Classification and nomenclature of viruses. 4th Report of the International Committee on Taxonomy of Viruses. Intervirology 1982;17:1-199.
- 2 Brown F: The classification and nomenclature of viruses: Summary of results of meetings of the International Committee on Taxonomy of Viruses, Edmonton, Canada, 1987. Intervirology 1989;30:181-186.
- 3 Atherton JG, Holmes IR, Jobbins EH: ICTV Code for the Decription of Virus Characters. Monogr Virol. Basel, Karger, 1983, vol 14.
- 4 Karabatsos N: International Catalogue of Arboviruses including Certain Other Viruses of Vertebrates. San Antonio, American Society of Tropical Medicine and Hygiene, 1985.

- 5 Jones E: Using dBASE III PLUS. Berkeley, Osborne McGraw-Hill, 1987.
- 6 Mueller J: Illustrated Clipper 5.0, New Delhi, BPB Publishers, 1991.
- 7 Blaine L: CODATA/IUIS Hybridoma Data Bank. Codata Bull 1991; 23:92-93.
- 8 George DG, Hunt LT, Barker WC: The National Biomedical Research Foundation Protein Sequence Database; in Lesk AM (ed): Computational Molecular Biology. Oxford, Oxford University Press, 1988, pp 17-26.
- 9 Nermut MV, Hockley DJ, Gelderblom H: Methods for the study of virus structure; in Nermut MV, Stevens A (eds): Perspectives in Medical Virology: Animal Virus Structure. Amsterdam, 1987, vol 3, pp 21-60.

- 10 Crowther RA: Procedures for three dimensional reconstruction of spherical viruses by Fourier synthesis from electron micrographs. Phil Trans R Soc Lond B 1971;261: 221–230.
- 11 Rogosa M, Krichevsky MI, Colwell RR: Coding Microbiological Data for Computers. New York, Springer, 1986.
- 12 Portyrata DA, Krichevsky MI: MICRO-IS: A microbiological database management and analysis system. Binary 1992;4:31–36.
- 13 Simons JF, Hellman U, Pettersson RF: Uukuniemi virus S RNA segment: Ambisense coding strategy, packaging of complementary strands into virions, and homology to members of the genus Phlebovirus. J Virol 1990;64:247–255.